

SPECIFICATION

SUPPORT, PRINTED MATERIAL AND REAGENT KIT HAVING ENZYME FIXED THEREON,
METHOD FOR PREPARING THE SUPPORT, METHOD FOR STORING ENZYME,
AND METHOD FOR RESTORATION ENZYMES

Field of the Invention

The present invention relates to a support, a printed material and a reagent kit having an enzyme fixed thereon, a method for preparing the support, a method for storing an enzyme, and a method for restoring an enzyme.

Background Art

Enzymes are proteins having catalytic activities and are involved in various biological reactions. Thus, enzymes contribute to the maintenance of life.

Enzymes are unstable at room temperature in the presence of moisture. Hence, enzymes are stored in a freeze-dried form or stored in a liquid containing a stabilizing agent at temperatures of -20°C or below.

Polymerase chain reaction (PCR) is a reaction for the amplification of a nucleic acid, which is catalyzed by an enzyme called "a DNA polymerase". A DNA polymerase is usually stored in a buffer at a temperature of -20°C . For this storage, a freezer is needed. When the enzyme is delivered from a supplier to a user, it is packaged in a container such as a container made of foamed polystyrene together with dry ice. These storage/delivery systems require specialized facilities and operations and, therefore, they are complicated and costly.

Disclosure of the Invention

Problems to be Resolved by the Invention

The object of the present invention is to provide a simple method

for storing an enzyme.

Means for Resolving the Problems

The present inventors have found that, when a DNA polymerase is fixed on a support as a mixture with trehalose for storage purpose and PCR is then performed using the support, the PCR reaction successfully proceeds. Based on this finding, the present invention has been accomplished.

The subject matters of the present invention are as follows.

(1) A support comprising an enzyme and a protecting agent for the enzyme fixed thereon.

(2) The support according to item 1, wherein the protecting agent is at least one chemical compound selected from the group consisting of trehalose and derivatives thereof.

(3) The support according to items 1 or 2, further comprising an enhancer for enzymatic reaction.

(4) The support according to any one of items 1 to 3, further comprising an aptamer for the enzyme thereon.

(5) A support comprising an enzyme and an aptamer for the enzyme fixed thereon.

(6) The support according to any one of items 1 to 5, wherein the enzyme is a DNA polymerase.

(7) The support according to item 6, further comprising a primer for the amplification of a nucleic acid of interest by a nucleic acid amplification reaction using the DNA polymerase.

(8) The support according to item 6, further comprising at least one member selected from the group consisting of a nucleic acid which serves as a template for the nucleic acid amplification reaction using the DNA polymerase, a primer for the amplification of the nucleic acid, and a buffer for the nucleic acid amplification reaction.

(9) A printed material comprising a support as recited in any one of items 1 to 8.

(10) A reagent kit comprising a support as recited in any one of items 1 to 8.

(11) A method for preparation of a support as recited in item 1, comprising: preparing a mixed solution of an enzyme and a protecting agent for the enzyme; applying the solution onto a support; and drying the support to fix a mixture of the enzyme and the protecting agent on the support.

(12) A method for restoration of an enzyme fixed on a support, comprising: immersing a support as recited in any one of items 1 to 8 in a liquid to leach out the enzyme into the liquid.

(13) A method for amplification of a nucleic acid, comprising: placing a support as recited in any one of items 6 to 8 in a liquid to leach out a DNA polymerase from the support; and performing a nucleic acid amplification reaction using the DNA polymerase.

The present invention also provides a method for storage of an enzyme in the form of being fixed on a support as a mixture with a protecting agent for the enzyme.

The present invention also provides a method for amplification of a nucleic acid, comprising: placing a support as recited in item (7) in a liquid to leach out a DNA polymerase and at least one member selected from the group consisting of a nucleic acid which serves as a template, primers for amplifying the nucleic acid and a buffer for the amplification reaction for the nucleic acid from the support; and performing the nucleic acid amplification reaction using the DNA polymerase and the nucleic acid which serves as a template and/or the primers.

Hereinbelow, the present invention will be described in detail.

The present invention provides a support having an enzyme and a protecting agent for the enzyme fixed thereon.

The enzyme may be any one as long as it has any catalytic activity. Examples of the enzyme include, but are not limited to, a DNA polymerase, an RNA polymerase, a reverse transcriptase, an RNase, a restriction enzyme,

a methylase, a modifying enzyme, a ligase, a protease, a kinase, a phosphatase, a transferase, a glycosylase, a topoisomerase and a clonase.

The protecting agent may be any one as long as it can protect an enzyme from drying and store the enzyme stably. Examples of the protecting agent include trehalose and derivatives thereof, polysaccharides, PEG, dextran, Ficoll, glycerol, surfactants, and PVA and derivatives thereof. Trehalose and derivatives thereof are particularly effective.

The protecting agent may be a commercially available product or may be synthesized according to any known method.

Trehalose is a non-reductive disaccharide composed of two 1,1-bonded D-glucose molecules and has three types of isomers: α,α -, α,β - and β,β -isomers, depending on the bonding manner.

Examples of the derivatives of trehalose include, but are not limited to, acid esters (e.g., fatty acid esters such as laurate, oleate, linoleate, linolenate, stearate, palmitate and myristate; carboxylates such as acetate and benzoate; and sulfate); alkyl ethers (e.g., ethers with C₈₋₂₅ alkyls); halides, nitrogen-containing derivatives and sulfur-containing derivatives of trehalose.

Trehalose and derivatives thereof are commercially available, but may be produced by any known method. The methods for production of trehalose and derivatives thereof can be found in, for example, "Developments in Food Carbohydrates", edited by C. K. Lee, issued by Applied Science Publishers, pp. 1-89, 1980; "Chemical and Pharmaceutical Bulletin", K. Yoshimoto et. al., vol. 30, No. 4, pp. 1,169-1,174, 1982; and Japanese Patent Application Laid-open No. 8-157491.

The protecting agent may be mixed with the enzyme in an amount of 10^{-5} to 10^1 M/U enzyme, preferably 10^{-4} to 10^{-1} M/U enzyme.

On the support, an enhancer for enzymatic reaction may also be fixed. The enzymatic reaction enhancer may be any substance as long as it has an effect of enhancing the desired enzymatic reaction. The effect of enhancing the desired enzymatic reaction includes an effect of preventing

the inhibition of the desired enzymatic reaction. Examples of the enzymatic reaction enhancer include, but are not limited to, sodium oxalate, potassium oxalate, sodium malonate, sodium maleate, dimethyl sulfoxide, betaines, glycerol, albumin, surfactants (e.g., Tween 20, Triton X100 and NP40), polyamines (e.g., ethylenediamine, trimethylenediamine, spermine, spermidine, diethylenetriamine, triethylenetetramine, tetraethylenepentamine, pentaethylenehexamine, 1,4-bis(3-aminopropyl)-piperazine, 1-(2-aminoethyl)piperazine, 1-(2-aminoethyl)piperidine, 1,4,10,13-tetraoxa-7,16-diazacyclooctadecane and tris(2-aminoethyl)amine), saccharides (e.g., glucose, fructose, galactose, maltose, sucrose, lactose and polysaccharides), sulfated polysaccharides and salts thereof (e.g., heparin, dextran sulfate), dithiothreitol, polyanions (e.g., DNA, RNA), polyhydric alcohols (e.g., aliphatic polyhydric alcohols such as ethylene glycol, propylene glycol, butanediol, hexanediol, octanediol, glycerin, sorbitan, trimethylol propane and neopentylglycol, diethylene glycol, triethylene glycol, polyethylene glycol and polypropylene glycol), ammonium sulfate and quaternary ammonium salts (e.g., hexadecyltrimethylammonium bromide, hexadecylpyridinium chloride, hexadimethrine bromide, hexafluorenum bromide and methylazolinium bromide). The enzymatic reaction enhancer includes, for example, Ampdirect® (Shimadzu Corporation), which is effective for enhancement of reactions with a DNA polymerase.

The enzymatic reaction enhancer may be fixed on the support in an appropriate amount. For example, a polyamine may be fixed on the support so that the polyamine is present in a reaction solution at a concentration of about 10 to 0.01 mM, preferably 2 to 0.5 mM. The enzymatic reaction enhancer may be fixed on the support in the same or different position as or from that of the mixture of the enzyme and the protecting agent.

The support may be any one as long as it can fix thereon the mixture of the enzyme and the protecting agent. Examples of the support include,

but are not limited to, paper such as 60MDP paper (a product by Mishima Paper Co., Ltd., Japan), copy paper, woodfree paper, mechanical paper, kent paper, drawing paper, craft paper, paper for inkjet printing, tracing paper, Japanese paper, board paper, filter paper; glass substrates; silicon substrates; beads; column fillers; silica gel; nitrocellulose membrane; nylon membrane; and PVA membrane.

The support may have a thickness of 1 mm or less, for example. With a very small thickness (for example, about 0.1 mm), the workability of the support can be improved even if a number of enzyme- and protecting agent-fixed supports are stacked for distribution purposes, because the supports are not so bulky.

In addition to the enzyme and the protecting agent, the support may further comprise other components fixed thereon, such as a polynucleotide (e.g., DNA, RNA, a derivative or modified form thereof), an oligonucleotide (e.g., DNA, RNA, a derivative or modified form thereof), a protein (e.g., an antibody, a hormone), a polypeptide, an oligopeptide, a polysaccharide, an oligosaccharide, PNA, a low molecular compound (e.g., EDTA, a salt contained in a PCR buffer composition, Mg^{2+} , a dNTP mixture) and a mixture thereof. The components other than the enzyme and the protecting agent may be fixed on the support in the same or different position as or from that of the mixture of the enzyme and the protecting agent. In particular, it is preferred that an aptamer for the enzyme be also fixed on the support. Accordingly, the present invention provides a support comprising an enzyme and an aptamer for the enzyme fixed thereon.

In one preferred embodiment of the present invention, in addition to the DNA polymerase and the protecting agent for the DNA polymerase, the support may further comprise primers for amplifying a nucleic acid of interest by nucleic acid amplification reaction using the DNA polymerase. The support of this type can be used in genotyping and identification of species. The support may further comprise an enhancer for the enzymatic reaction.

In another preferred embodiment of the present invention, in addition to the DNA polymerase and the protecting agent for the DNA polymerase, the support may further comprise at least one member selected from the group consisting of a nucleic acid which serves as a template for a nucleic acid amplification reaction (e.g., PCR) using the DNA polymerase, a primer for amplifying the nucleic acid, and a buffer for the nucleic acid amplification reaction. The support may also comprise an enhancer for enzymatic reaction.

For example, when a DNA polymerase is fixed on paper (support) for storage purpose, in addition to the DNA polymerase and the protecting agent, the paper may further have a set of primers (oligonucleotides), DNA which serves as a template for PCR reaction (which may be synthetic single- or double-stranded DNA or a vector having cDNA cloned therein), an aptamer for the DNA polymerase (functional RNA), components to be contained in a PCR reaction solution (i.e., Tris-HCl, KCl, MgCl, a dNTP mixture, etc.), EDTA and the like fixed thereon. In this case, fixation of these components may be achieved by any one of the following procedures: (1) the DNA polymerase, the protecting agent and the primer set are fixed together on the paper as a single spot, and the DNA as a template for PCR reaction, Tris-HCl and EDTA are fixed together on the paper as a separate single spot; (2) the DNA polymerase, the protecting agent, the primer set and, if required, an aptamer for the DNA polymerase are fixed together on the paper as a single spot; or (3) all of the components required for PCR reaction (i.e., DNA as a template for PCR reaction, the DNA polymerase, the primer set, Tris-HCl, KCl, MgCl, a dNTP mixture and the like and, if required, an aptamer for the DNA polymerase) together with the protecting agent are fixed on the paper as a single spot. For ease of distinction of the component (e.g., a DNA polymerase) spotted on paper, the component to be spotted may be mixed with a dye. Examples of the dye include, but are not limited to, cresol red, bromophenol blue and xylene cyanol.

The amount of the enzyme to be fixed on the support may be properly selected so that the desired enzymatic reaction can be achieved. For example, for PCR reaction, 5 ng or more of a DNA polymerase may be fixed per spot.

A support having a mixture of an enzyme and a protecting agent for the enzyme fixed thereon can be prepared in the following manner. First, a mixed solution of an enzyme and a protecting agent is prepared. The mixing ratio between the enzyme and the protecting agent is as described above. The solvent is preferably water. The mixed solution may further contain any one of the components other than the enzyme and the protecting agent as described above. Next, the mixed solution of the enzyme and the protecting agent is applied onto a support. For example, in the case where the support is paper, the mixed solution can be spotted on the paper using a syringe, a 96 pin-tool (Multi 96-multiblot replicator VP409, Bio Medical Science Inc., US), a disposable-type pin-tool or the like. The support is then dried to fix the mixture of the enzyme and the protecting agent thereon. Preferably, the support having the mixture of the enzyme and the protecting agent fixed thereon contains substantially no moisture.

By fixing an enzyme on a support as a mixture with a protecting agent for the enzyme as described above, the enzyme can be stored stably. As for the storage conditions, the support is preferably stored at room temperature under light shielding conditions while avoiding high humidities. In the case where the enzyme is a DNA polymerase, for example, when the DNA polymerase is stored at room temperature after being fixed on 60MDP paper as a mixture with trehalose, storage life of at least 6.5 months has been confirmed (the storage test is now being carried on).

For restoration of an enzyme fixed on a support as a mixture with a protecting agent for the enzyme as described above, the support having the mixture of the enzyme and the protecting agent fixed thereon may be immersed in a liquid to leach out the enzyme into the liquid. The liquid to be used for the immersion of the support may be any one as long as

it enables the restoration of the enzyme. Examples of the liquid includes, but are not limited to, water and an aqueous solution containing an ingredient other than water. In the case where the enzyme fixed on a support is a DNA polymerase, for example, the liquid to be used for the immersion of the support is preferably water, a PCR reaction solution (i.e., an aqueous solution containing Tris-HCl, KCl, MgCl, a dNTP mixture and the like) or the like. The immersion may be performed at room temperature under atmospheric pressure for 1 to 3 minutes.

When a DNA polymerase is fixed on a support, the support may be placed in a liquid to leach out the DNA polymerase from the support and nucleic acid amplification reaction may be then performed using the leached-out DNA polymerase to amplify a nucleic acid.

The present invention also provides a printed material comprising a support having an enzyme and a protecting agent for the enzyme fixed thereon.

The printed material includes, but is not limited to, complete books (e.g., textbooks), handbooks, catalogues, journals, magazines, articles, , booklets, minibooklets, leaflets, pamphlets, reports, posters, cards and labels.

Fig. 1 shows an embodiment of the printed material according to the present invention, in which a mixture of an enzyme (a DNA polymerase) and trehalose together with a primer set, a cDNA clone which serves as a template for PCR reaction and other components required for the PCR reaction (i.e., Tris-HCl, KCl, MgCl, each dNTP and, if required, an aptamer for the DNA polymerase) is spotted on paper (support). The DNA polymerase and trehalose are spotted on the paper 6 (hereinafter, this spot is referred to as "DNA Spot 1"). In the paper 6, in addition to DNA spot 1, the name 2 of a protein encoded by the cDNA clone ("malate dehydrogenase"), the identification number 3 of the cDNA clone ("Clone ID"), the nucleotide sequence 4 of the cDNA clone ("DNA sequence") and the instructions 5 of the procedure for an experiment (PCR reaction)

("Procedures") are printed on the paper.

Fig. 2 shows a magazine 13 including a scientific article 12, to which the paper 6 having DNA spot 1 thereon shown in Fig. 1 is added as an appendix.

Fig. 3 shows a book 22 in which the paper 6 having DNA spot 1 thereon shown in Fig. 1 is bounded. This book may further include a table of contents.

Fig. 4 shows another embodiment of the book in which paper (support) comprising a mixture of an enzyme (a DNA polymerase) and trehalose spotted thereon is bounded. On each grid of a checkerboard pattern of page 34 on which a mixture of an enzyme (a DNA polymerase) and trehalose are spotted, the DNA polymerase and trehalose together with a primer set, a cDNA clone which serves as a template for PCR reaction and other components required for the PCR reaction (i.e., Tris-HCl, KCl, MgCl, each dNTP and, if required, an aptamer for the DNA polymerase) are spotted (hereinafter, this spot is referred to as "DNA spot 31"). On the page 34 having these spots 31, symbols 32 (column number) and 33 (row number) for identification of each spot are printed. Also printed are the identification number 30 ("Rearray PLATE ID") of a page having DNA spotted thereon. The information about the spotted cDNA clone, such as EC number of an enzyme encoded by the cDNA clone, name of an enzyme encoded by the cDNA clone ("Gene name"), ID number ("RIKEN Clone ID") and deposition number ("Accession Number") of the clone, length of the cDNA clone insert ("cDNA Insert), length of a PCR reaction product ("After PCR") and the explanation of a reaction in which the enzyme encoded by the cDNA clone is involved, and the information about the primer set, such as nucleotide sequence of the primers, are recorded in a CD-ROM 36 (as a substitute for the CD-ROM, other recording medium such as FD and MD may be used). The recording medium is added as an appendix to the book (Fig. 5). In Fig. 5, the CD-ROM 36 is packed in a bag 37 and attached to the book 35 with the bag 37 being sealed with a seal 38. This book may also have a page on which a table

of contents, an instruction manual for the spots containing the cDNA clone and primer set, and an access guide to the information recorded in the recording medium are printed.

Exemplary types of the printed material include the following: 1) an encyclopedia-like all-inclusive type (e.g., FANTOM clone, human metabolome); 2) a volume-separated type by item (e.g., function, organ); 3) a small volume type by subdivided subjects or contents which consists of one to several pages (e.g., loose-leaf type); and 4) a card type which is intended to have a smaller number of attachments.

The present invention also provides a reagent kit comprising a support having an enzyme and a protecting agent for the enzyme fixed thereon.

The reagent kit according to the present invention can be used as a nucleic acid amplification reaction (e.g., PCR) kit, a protein production kit, an antibody kit and other types of kits for use in a variety of experiments, tests, diagnoses and the like.

The reagent kit according to the present invention may be in the form of a printed material as described above. Other embodiments of the reagent kit are shown in Figs. 6 to 9.

Fig. 6 shows an embodiment of the reagent kit according to the present invention, in which a mixture of an enzyme (a DNA polymerase) and trehalose together with a primer set and other components required for PCR reaction (i.e., Tris-HCl, KCl, MgCl, each dNTP and, if required, an aptamer for the DNA polymerase) is spotted on paper (support). In the paper, the DNA polymerase and trehalose are spotted in a proper position on the paper (hereinafter, this spot is referred to as "DNA spot 41").

Fig. 7 shows an embodiment of the reagent kit, which comprises a sheet of paper 42 having a DNA spot 41 shown in Fig. 6 thereon. The paper 42 having the DNA spot 41 is placed in a light-proof bottle 51, and then stored or distributed with the bottle 51 hermetically sealed with a cap 52. The reagent kit may further include an instruction manual 53 on which

the information about the kit, such as the contents of the kit (e.g., names/quantities of components included in the kit, recommended use, directions for storage/expiration date, package unit), directions for use, cautions in use and handling, and customer inquiry, is printed. The instruction manual 53 may be included in the light-proof bottle 51 or placed in a package box (now shown) including the light-proof bottle 51. Alternatively, the instruction manual may be printed on a label and attached on the light-proof bottle 51.

Fig. 8 shows an embodiment of the reagent kit according to the present invention, in which a mixture of an enzyme (a DNA polymerase) and trehalose is spotted on paper (support). In the paper 62, the DNA polymerase and trehalose together with a primer set and other components required for PCR reaction (i.e., Tris-HCl, KCl, MgCl, each dNTP and, if required, an aptamer for the DNA polymerase) are spotted in a proper position on the paper (hereinafter, this spot is referred to as "DNA spot 61").

Fig. 9 shows an embodiment of the reagent kit comprising a sheet of paper 62 having DNA spots 61 thereon. The paper 62 having DNA spots 61 is placed in a packaging pack 71 and stored or distributed with the packaging pack 71 being hermetically sealed. The reagent kit may further include an instruction manual 72 on which the information about the kit, such as the contents of the kit (e.g., names/quantities of components included in the kit, recommended use, directions for storage/expiration date, package unit), directions for use, cautions in use and handling, and customer inquiry, is printed. The instruction manual 72 may be included in the packaging pack 71 or placed in a package box (now shown) including the packaging pack 71. Alternatively, the instruction manual 72 may be printed on a label and attached on the packaging pack 71 or the package box.

Hereinabove, the present invention is described with reference to several embodiments of the combination of a DNA polymerase and DNA. However, the invention is not limited to these embodiments and applicable

to a variety of enzymes.

In this specification, it is noted that the mark "--" means a numerical range in which the numerical numbers given before and after the mark are included in the range as the minimum value and the maximum value, respectively.

Effect of the Invention

According to the present invention, a simple method for storing an enzyme is provided.

This specification includes part or all of the contents as disclosed in the specification and/or drawings of Japanese Patent Application No. 2003-339542 based on which the present application claims priority.

Brief Description of Drawings

Fig. 1 shows an embodiment of paper (support) comprising a mixture of a DNA polymerase and trehalose together with DNA (cDNA, primer, aptamer, etc.) spotted thereon.

Fig. 2 shows an embodiment of a magazine including a scientific article, in which paper comprising a mixture of a DNA polymerase and trehalose together with DNA (cDNA, primer, aptamer, etc.) spotted thereon is added as an appendix.

Fig. 3 shows an embodiment of a book in which paper comprising a mixture of a DNA polymerase and trehalose together with DNA (cDNA, primer, aptamer, etc.) spotted thereon is bounded.

Fig. 4 shows another embodiment of a book in which paper (support) comprising a mixture of a DNA polymerase and trehalose together with DNA (cDNA, primer, aptamer, etc.) spotted thereon is bounded.

Fig. 5 shows an embodiment of packaging into a bag a CD-ROM on which the information about cDNA spotted on the paper shown in Fig. 4 is recorded, with the CD-ROM being added as an appendix to the book with the bag sealed with a seal.

Fig. 6 shows an embodiment of paper (support) comprising a mixture of a DNA polymerase and trehalose together with DNA (cDNA, primer, aptamer, etc.) spotted thereon.

Fig. 7 shows an embodiment of a reagent kit comprising the paper shown in Fig. 6.

Fig. 8 shows an embodiment of paper (support) comprising a mixture of a DNA polymerase and trehalose together with DNA (cDNA, primer, aptamer, etc.) spotted thereon.

Fig. 9 shows an embodiment of a reagent kit comprising the paper shown in Fig. 8.

Fig. 10 shows the constitution of pFLC vector in which murine malate dehydrogenase cDNA is cloned.

Fig. 11 shows 60MDP paper having both a spot of a solution of murine malate dehydrogenase cDNA and a spot of a [polymerase + primer] solution.

Fig. 12 shows the electrophoretic results of a product of the PCR reaction performed using the spots shown in Fig. 11.

Fig. 13 shows 60MDP paper having a spot of a [primer + aptamer + polymerase] solution.

Fig. 14 shows the electrophoretic results of a product of the PCR reaction performed using the spot shown in Fig. 13.

Fig. 15 shows 60MDP paper having spots of a [cDNA + primer + aptamer + polymerase + PCR buffer composition] solution, wherein the cDNA is that for any one of murine malate dehydrogenase, murine isocitrate dehydrogenase (NADP), murine isocitrate dehydrogenase (NAD) and murine oxoglutarate dehydrogenase.

Fig. 16 shows the electrophoretic results of a product of the PCR reaction performed using the spots shown in Fig. 15.

Fig. 17 shows the electrophoretic results of a product of the PCR reaction performed using the spots of a [cDNA + primer + reaction enhancer (spermidine) + polymerase + PCR buffer composition] solution.

Explanation of Reference Signs in the Drawings

- 1: DNA spots
- 2: Name of protein (enzyme such as malate dehydrogenase) encoded by cDNA clone
- 3: Identification number of cDNA clone
- 4: Nucleotide sequence of cDNA clone
- 5: Instruction manual for experimental procedure
- 6: Paper having a mixture of DNA polymerase and trehalose together with DNA (cDNA, primer, aptamer, etc.) spotted thereon
- 12: Scientific article
- 13: Magazine
- 22: Book
- 30: Identification number of page on which DNA is spotted
- 31: DNA spots
- 32: Symbol 1 for identification of spot (column number)
- 33: Symbol 2 for identification of spot (row number)
- 34: Page having a mixture of DNA polymerase and trehalose together with DNA (cDNA, primer, aptamer, etc.) spotted thereon
- 35: Book
- 36 CD-ROM
- 37: Bag
- 38: Seal
- 41: DNA spot
- 42: Paper (support)
- 51: Light-proof bottle
- 52: Cap
- 53: Instruction manual
- 61: DNA spots
- 62: Paper (support)
- 71: Packaging pack
- 72: Instruction manual

Best Modes for Carrying out the Invention

Hereinbelow, the present invention will be described in great detail with reference to the following examples. However, the examples are illustrative only and the scope of the invention is not limited to these examples.

Examples

[Example 1]

DNA bank having spots of cDNA clone and polymerase thereon

<Synthesis of primers>

A set of primers having the following sequences were synthesized according to a conventional method.

Primer set 1:

-21M13: 5'-TGTAACGACGGCCAGT-3' (SEQ ID NO:1)

1233-Rv: 5'-AGCGGATAACAATTTACACAGGA-3' (SEQ ID NO:2)

<Preparation of cDNA solution>

pFLC vector, into which murine malate dehydrogenase cDNA (Clone ID: 1500012M15, 1758 bp) having the following nucleotide sequence collected from the Riken clones (<http://fantom.gsc.riken.go.jp/>) had been cloned (Fig. 10), was dissolved in TE buffer (10 mM Tris-HCl, pH 8.0, 1 mM EDTA) to give a concentration of 0.1 µg/µl.

Murine malate dehydrogenase 1500012M15

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1  cccggttctc tcccagagtc tgttccgctg tagaggtgac ctgactgctg gagactgcct
61  tttgcagggtg cagagatcgg ccttgcagtt tgcaataatg tctgaaccaa tcagagtcct
121 tgtgactgga gcagctggtc aaattgcata ttcactgttg tacagtattg gaaatggatc
181 tgtctttggg aaagaccagc ccatcattct tgtgctgttg gacatcaccc ccatgatggg
241 tgttctggac ggtgtcctga tggaactgca agactgtgcc cttccccttc tgcaggatgt
301 cattgcaacg gacaaagaag agattgcctt caaagacctg gatgtggctg tcctagtggg
361 ctccatgcca ataaggggaag gcatggagag gaaggacctc ctgaaagcca atgtgaaaat
421 cttcaaatac cagggcacag ccttggagaa atacgccaag aatcagtta aggtcattgt
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481 tgtgggaaac ccagccaata cgaactgcct gacagcctcc aagtcagcgc catcgatccc
 541 caaggagaat ttcagttgcc tgactcgctt ggaccacaac cgagcaaaat ctcaaattgc
 601 tcttaaactc ggtgtaaccg ctgatgatgt aaagaatgtc attatctggg gaaatcattc
 661 atcgaccag tatccagatg tcaatcatgc caaggtgaaa ctgcaaggaa aggaagtcgg
 721 tgtgtatgaa gccctgaaag acgacagctg gctgaaggga gagttcatca cgactgtgca
 781 acagcgtggt gctgctgtca tcaaggctcg gaagctgtcc agtgcaatgt ctgctgcgaa
 841 agccatcgca gaccacatca gagacatctg gtttggaacc ccagagggag agttcgtgtc
 901 gatgggtggt atctctgatg gcaactccta tgggtgtccct gatgacctgc tctactcatt
 961 ccctgtcgtg atcaagaata agacctggaa gtttggtgaa ggcctcccca ttaatgactt
 1021 ctcccgtgaa aagatggacc tgacagcaaa ggagctgacc gaggaaaagg agaccgcttt
 1081 tgagtttctc tcctctgcgt gactagacac tcgttttgac atcagcagac agccgaaggc
 1141 tgaggaatca aaatgtcgtc tttgagccta gtaccaaaca gtaataatgc tacattcaaa
 1201 ttgtgaacag caaaatattt taaatagtgt gtgctttatg atttgtgaaa gtctatcatg
 1261 ttgttagtgc tgcaatctaa ataaaagtat attcaagtga aaatctctca gactctgttt
 1321 ctactttata tttagtatct tcaggaaaac aagtttgccc aatagattat aattttactt
 1381 ttttaattga ctaaaagaaa taaagatgga aaatattatg aagtaaagca ttagtctcta
 1441 acataaacia ggaagcccaa tcaatttcag agggatccca ttacttaagt ccttaaaggc
 1501 tggttcatgt tttgctcata atttgatttt aaaattagct gtaagaaggc tgcagataat
 1561 ctatcttctt tatattctat agcagaataa tgaagtcatt aatatttgat agccaataat
 1621 accacactat taatatttgt aagctaagat tattagaaac ataaaactgt ttttgagtca
 1681 gtctgttttc catgagaaga catgcatcat ctttgtgtgt tttgtgcatt actcagtgc
 1741 ataaataacc ataattctc (SEQ ID NO:3)

<Preparation of [polymerase + primer] solution>

KOD plus polymerase (Toyobo Engineering Co., Ltd., Japan), trehalose and the primer set 1 were mixed together to prepare a solution having the final concentrations of 25 U/ μ l of KOD plus DNA polymerase, 0.1 M of trehalose and 2 μ M of primer set 1.

<Spotting of DNA>

The cDNA solution and the [polymerase + primer] solution were spotted on a sheet of 60MDP paper (Mishima Paper Co., Ltd., Japan) using a 96 pin-tool (Multi 96-multiblot replicator VP409, Bio Medical Science

Inc., US) so that the positions and the types of the spots could be identified, as shown in Fig. 11. The cDNA solution and the [polymerase + primer] solution were spotted at rates of 0.5 μ l/spot and 1 μ l/spot, respectively.

<Recovery and amplification of DNA>

The spotted paper was dried at room temperature for at least 30 minutes. Thereafter, two 4 mm x 4 mm pieces were cut out from the 60MDP paper so that each piece contained the spotted cDNA or [polymerase + primer] and then placed in a PCR microtube. The tube was added with 25 μ l of a PCR reaction solution (10 mM Tris-HCl, pH 8.3, 50 mM KCl, 5.3 mM MgCl₂, 200 μ M each dNTP) and PCR was run under the following conditions:

2 min at 94°C;

29 cycles of (1 min at 94°C, 1 min at 55°C, 75 sec at 68°C); and
then

15 min at 74°C.

After the reaction was completed, a proper amount of the reaction solution was removed from the tube and then subjected to 1% agarose gel electrophoresis. The results are shown in Fig. 12. A band observed at the position around 1800 bp was considered to be the desired fragment. It was demonstrated that the DNA was leached out from the 60MDP paper and could be amplified by PCR.

[Example 2]

Experiment of [aptamer + polymerase] using lutenizing hormone gene

<Synthesis of primers>

Primers having the following sequences were synthesized according to a conventional method.

Primer set 1 (a primer set for amplification of human lutenizing hormone gene exon 1):

HsLH1F: CCAGGGGCTGCTGCTGTTG (SEQ ID NO:4)

HsLH1R: CATGGTGGGGCAGTAGCC (SEQ ID NO:5)

Primer sets 2 (a primer set for amplification of human lutenizing hormone gene exon 2):

HsLH2F: ATGCGCGTGCTGCAGGCG (SEQ ID NO:6)

HsLH2R: TCGGGATTGAGAAGCCTTTATTG (SEQ ID NO:7)

<Synthesis of aptamer>

An oligonucleotide having the following sequence, which is a known aptamer for Taq DNA polymerase (Yun Lin, Sumedha D. Jayasena, Inhibition of Multiple Thermostable DNA Polymerases by a Heterodimeric Aptamer, Journal of Molecular Biology (1997), Vol. 27, Issue 1, pages 100-11), was synthesized by a conventional method.

GCCGGCCAATGTACAGTATTGGCCGGC (SEQ ID NO:8)

<Preparation of [primer + aptamer + polymerase] solution>

Two kinds of spotting solutions were prepared.

The above primer sets, the aptamer for Taq DNA polymerase and Taq DNA polymerase were mixed together to prepare a solution having the final concentrations of 2 μ M of the primer sets, 2 μ M of the aptamer for Taq DNA polymerase, 25 U/ μ l of Taq DNA polymerase and 0.1 M trehalose. Another solution having the same composition except that the aptamer for Taq DNA polymerase was eliminated was also prepared.

<Spotting of DNA>

Each of the [primer + aptamer + polymerase] solutions prepared as described above was spotted on a sheet of 60MDP paper (Mishima Paper Co., Ltd., Japan) using a 96 pin-tool (Multi 96-multiblot replicator VP409, Bio Medical Science Inc., US) so that the position and the type of the spot could be identified, as shown in Fig. 13. The spotting solution was applied at a rate of 1 μ l/spot.

<Amplification of DNA>

The spotted paper was dried at room temperature for at least 30 minutes. Thereafter, a 4 mm x 4 mm piece was cut out from the 60MDP paper so that the piece contained the spotted area and then placed in a PCR microtube. The tube was added with 25 μ l of a PCR reaction solution (10

mM Tris-HCl, pH 8.3, 50 mM KCl, 5.3 mM MgCl, 200 μ M each dNTP) and 50 ng of template DNA (human genomic DNA; BD Biosciences Clontech, US), and PCR was run under the following conditions:

3 min at 94°C;

50 cycles of (30 sec at 94°C, 30 sec at 40°C, 30 sec at 72°C); and then

15 min at 72°C.

After the reaction was completed, a proper amount of the reaction solution was removed from the tube and then subjected to 1% agarose gel electrophoresis. The results are shown in Fig. 14. Bands observed at the positions around 184 bp and 343 bp were considered to be the desired DNA fragments of exons 1 and 2, respectively. It was demonstrated that fragments of interest could be amplified by PCR from the template DNA by using the primers fixed on the 60MDP paper. Comparison was made between the samples with and without the aptamer for Taq DNA polymerase, and it was found that the reaction with the aptamer could inhibit non-specific amplification.

[Example 3]

DNA bank having spots of Riken cDNA clone + PCR solution thereon

<Synthesis of primers>

A set of primers having the following sequences were synthesized according to a conventional method.

Primer set 1:

-21M13: 5'-TGTAACGACGGCCAGT-3' (SEQ ID: NO:1)

1233-Rv: 5'-AGCGGATAACAATTTACACAGGA-3' (SEQ ID NO:2)

<Preparation of cDNA solution>

pFLC vector, into which any one of murine malate dehydrogenase cDNA (Clone ID: 1500012M15, 1758 bp), murine isocitrate dehydrogenase (NADP) (Clone ID: 1500012E04, 2440 bp), murine isocitrate dehydrogenase (NAD) (Clone ID: E030024J03, 2160 bp) and murine oxoglutarate dehydrogenase

(Clone ID: E430020N12, 3554 bp) having the following nucleotide sequences collected from the Riken clones (<http://fantom.gsc.riken.go.jp/>) had been cloned (Fig. 10), was dissolved in TE buffer (10 mM Tris-HCl, pH 8.0, 1 mM EDTA) to give a concentration of 0.1 µg/µl.

Murine malate dehydrogenase 1500012M15

```

1  cccggttctc tcccagagtc tgttccgctg tagaggtgac ctgactgctg gagactgcct
61  tttgcaggtg cagagatcgg ccttgcagtt tgcaataatg tctgaaccaa tcagagtcct
121 tgtgactgga gcagctggtc aaattgcata ttcactgttg tacagtattg gaaatggatc
181 tgtctttggg aaagaccagc ccatcattct tgtgctggtg gacatcaccc ccatgatggg
241 tgttctggac ggtgtcctga tggaaactgca agactgtgcc cttccccttc tgcaggatgt
301 cattgcaacg gacaaagaag agattgcctt caaagacctg gatgtggctg tcctagtggg
361 ctccatgcca ataaggggaag gcatggagag gaaggacctc ctgaaagcca atgtgaaaat
421 cttcaaatcc cagggcacag ccttggagaa atacgccaag aaatcagtta aggtcattgt
481 tgtgggaaac ccagccaata cgaactgcct gacagcctcc aagtcagcgc catcgatccc
541 caaggagaat ttcagttgcc tgactcgctt ggaccacaac cgagcaaaat ctcaaattgc
601 tcttaaactc ggtgtaaccg ctgatgatgt aaagaatgtc attatctggg gaaatcattc
661 atcgacccag tatccagatg tcaatcatgc caaggtgaaa ctgcaaggaa aggaagtcgg
721 tgtgtatgaa gccctgaaag acgacagctg gctgaaggga gagttcatca cgactgtgca
781 acagcgtggt gctgctgtca tcaaggctcg gaagctgtcc agtgcaatgt ctgctgcgaa
841 agccatcgca gaccacatca gagacatctg gtttggaacc ccagagggag agttcgtgtc
901 gatgggtggt atctctgatg gcaactccta tgggtgtcct gatgacctgc tctactcatt
961 ccctgtcgtg atcaagaata agacctggaa gtttggtgaa ggccctccca ttaatgactt
1021 ctcccgtgaa aagatggacc tgacagcaaa ggagctgacc gaggaaaagg agaccgcttt
1081 tgagtttctc tcctctgcgt gactagacac tcgttttgac atcagcagac agccgaaggc
1141 tgaggaatca aaatgtcgtc tttgagccta gtaccaaaca gtaataatgc tacattcaaa
1201 ttgtgaacag caaaatattt taaatagtgt gtgctttatg atttgtgaaa gtctatcatg
1261 ttgttagtgc tgcaatctaa ataaaagtat attcaagtga aaatctctca gactctgttt
1321 ctactttata tttagtatct tcaggaaaac aagtttgccc aatagattat aattttactt
1381 ttttaattga ctaaaagaaa taaagatgga aaatattatg aagtaaagca ttagtctcta
1441 acataaacia ggaagcccaa tcaatttcag aggatccca ttacttaagt ccttaaaggt

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1501 tggttcatgt tttgctcata atttgatttt aaaattagct gtaagaaggt tgcagataat
 1561 ctatcttctt tatattctat agcagaataa tgaagtcatt aatatttgat agccaataat
 1621 accacactat taatatttgt aagctaagat tattagaaac ataaaactgt ttttgagtca
 1681 gtctgttttc catgagaaga catgcatcat ctttgtgtgt tttgtgcatt actcagtgca
 1741 ataaataacc ataatctc (SEQ ID NO:3)

Murine isocitrate dehydrogenase (NADP) 1500012E04

1 ggggtgttgcc gctgtcgccg cggtgagga agtggacgcg atggccgggt ccgcgtgggt
 61 gtccaagggtc tctcggctgc tgggtgcatt ccacaacaca aaacagggtga caagagggtt
 121 tgctggtggt gttcagacag taactttaat tcctggagat ggaattggcc cagaaatttc
 181 agcctcagtc atgaagattt ttgatgctgg ccaaagcacc tattcagtgg gaggagcgca
 241 atgtcacagc aattcaagga ccaggaggaa agtgggatga tccctccaga agccaaggag
 301 tccatggata agaacaagat gggcttgaaa ggcccactaa agaccccaat agccgctggc
 361 catccatcta tgaatctgtt gcttcgtaag acatttgacc tttatgccaa tgtccggcca
 421 tgtgtctcaa ttgaaggta taaaaccctt tacacggatg taaatatcgt caccatccga
 481 gagaacacgg aaggagaata cagtgaatt gagcatgtga tcgttgatgg ggttgtgcag
 541 agcatcaagc tcatcaccga agaagcaagc aagcgcattg cagagtttgc cttcgagtac
 601 gctcgggaaca accaccggag caacgtcaca gctgtgcaca aagctaacat catgaggatg
 661 tcagatgggc tctttctgca aaaatgcagg gaagttgcgg agaactgtaa agacattaaa
 721 ttaacgaga tgtacctga tactgtatgt taaatatgg tacaagacc atccagttt
 781 gatgttcttg tcatgccaaa ttatacga gacatcctta gtgatctgtg tgcaggactg
 841 attggagggtc ttgggggtgac tccaagtggc aatattggag ccaacgggtg tgccatcttt
 901 gaatcggttc atggaacagc cccggacatt gcaggcaagg acatggccaa cccacggcc
 961 ctctgctta gtgctgtgat gatgcttcgc cacatgggac tttttgacca tgcagcaaaa
 1021 atcgaggctg catgttttgc tacaattaag gatggaaaga gcttaacaaa agatctggga
 1081 ggcaacgcga agtgctctga cttcacagaa gaaatctgtc gtagagtcaa agacttagat
 1141 tagcactcct gctggtggat ttgctgcagt cagtcaatca ctccaaaagg ataccctgta
 1201 atcctccttg agggcgccca ccattgggtt gcttgcttct tgacagagta cgttttttga
 1261 atctggcctt ttcttaacaa aacccttgca atggatgcac atgatggccc caggccttca
 1321 ttcaaagggt tttcccaagt gctgggttga tttattgtcc gtctggtaaa ccttattttg
 1381 taaactgtaa gtgaactgta tcatttatca ttgttaacct attttacact tcaggcaaaa

1441 tcatttttcct caactgtaaa tattctgata cagaattaat aagagaagat atttaacttt
 1501 ttaacaaaag ccctggattt ttgggtttatg aaaaacaaac tgggaataaa acaggggtttc
 1561 aacaatcgca caagataaca ttatttctaata actaatgggt acaaaagaaa ttactggga
 1621 aagttcacag caaaaaactg gtatatatttct taaaaatatg gaaataaagt atttgtccta
 1681 tacatgaatt actattaata aaaatgtaag ctccaagaaa tccataatga atgatgtaat
 1741 tttgttacta catcggtaat ccttgtcaag gccccggatg ctctctgtgt atttgattct
 1801 ttgggttacct tgagattcac tatttggggg gaagagcttt cagataaggg agatcactcc
 1861 tcactagaca gatcgtcagc attgcgagct gtcagccatg agagccagcc actgcagatc
 1921 ccctcccacg tggccacact ccagccagtg ctgcaggtga ccctggaaag gcctggctgc
 1981 cccttgactt tccctaaagc aaccagtcac tgccttctgc ccagtagca cccattacag
 2041 acttaattgc cgaggtggag ctgactcagc ccacgctcat acaaatcagg ccaagcgggg
 2101 gcctgtgtta ccagctgctg accatcaggt tctgcccctc attcttccca cagcctctgc
 2161 tccacagcat gaacctagcc tttggcccac accaaagcca agctgtcttc ccttagccct
 2221 tgcactagtt tgcaaactcg tggctttgca taatgtaccc tgggcccaag gggatttctt
 2281 aacaacagat gtccctgtct gggtcatttt tttaaagctt ttatttggac ttacaatctt
 2341 ctgtgtatatt tacttttaaaa ctgctgcttt ccctgtctca ctggattggt ctgggttagca
 2401 gtggctttgg gttcacagta ataaagaact taagaact (SEQ ID NO:9)

Murine isocitrate dehydrogenase (NAD) E030024J03

1 ggatctaact ggggccggct tattacagct tgtgtgtacg cgcggggtgtg agccgggtta
 61 ttgaagtaaa aatgtccaga aaaatccaag gaggttctgt ggtggagatg caaggagatg
 121 aaatgacacg aatcatttgg gaattgatta aggaaaaact tattcttccc tatgtggaac
 181 tggatctgca tagctatgat ttaggcatag agaatcgtga tgccaccaat gaccaggtca
 241 ccaaagatgc tgcagaggct ataaagaaat acaacgtggg cgtcaagtgt gctaccatca
 301 ccccgatga gaagagggtt gaagaattca agttgaaaca aatgtggaaa tccccaaatg
 361 gcaccatccg aaacattctg ggtggcactg tcttcaggga agctattatc tgcaaaaata
 421 tccccgggt agtgacaggc tgggtaaaac ccatcatcat tggccgacat gcatatgggg
 481 accaatacag agcaactgat tttgttggtc ctgggcctgg aaaagtagag ataacctaca
 541 caccaaaaga tggaactcag aaggtgacat acatggtaca tgactttgaa gaaggtgggtg
 601 gtgttgccat gggcatgtac aaccaggata agtcaattga agactttgca cacagttcct
 661 tccaaatggc tctgtccaag ggctggcctt tgtatctcag caccaagaac actattctga

721 agaagtatga tgggcggttc aaagacatct tccaggagat ctatgacaag aaatacaagt
 781 cccagtttga agctcagaag atctgctatg aacacaggct catagatgac atgggtggccc
 841 aagctatgaa gtccgaggga ggcttcatct gggcctgtaa gaattacgat ggggatgtgc
 901 agtcagactc agtcgcccaa ggttatggct cccttggcat gatgaccagt gtgctgattt
 961 gtccagatgg taagacggta gaagcagagg ctgcccatgg cactgtcaca cgtcactacc
 1021 gcatgtacca gaaagggcaa gagacgtcca ccaaccccat tgcttccatt tttgcctggg
 1081 cccgagggtt agcccacaga gcaaagcttg ataacaatac tgagctcagc ttcttcgcaa
 1141 aggctttgga agacgtctgc attgagacca ttgaggctgg ctttatgact aaggacttgg
 1201 ctgcttgcac taaaggctta cccaatgtac aacgttctga ctacttgaat acatttgagt
 1261 ttatggacaa acttgagaa aacttgaagg ccaaattagc tcaggcccaa actttaagg
 1321 caaacctggg cttagaatga gtctttgcgg taactaggct cacagggtta cgtatttttt
 1381 tttttttttt tagtaacact caagattaaa aacaaaaatc attttgtaat tggtttagaa
 1441 gacaaagttg aacttttata tatgtttaca gtcttttttc tttttcatac agttattgcc
 1501 acctaatga atgtgggtggg gaaatttttt taattgtatt ttattgtgta gtagcagtgt
 1561 aggaattatg ttagtacctg ttcacaatta actgtcatgt tttctcatgc tctaattgtaa
 1621 atgaccaaaa tcagaagtgc tccaagggtg aacaatagct acagtatggg tccccataag
 1681 gggaaaagag aaactcactt cccctgttgt ccatgagtgt gaacactggg gcctttgtac
 1741 gcaaatgttg tactgtgtgt gggagagcta tacagtaagc tcacataaga ctggaacaga
 1801 taggatgtgt gtagctaaaa tgcattggcag acgtgtttat aaagagcatg tatgtgtcca
 1861 atatactagt tatattttta gaccactgga gaattccaag tctagaataa atgcagactg
 1921 gaggattctg ctctttgatt tctcttctcc tgtgaccag cctaagtatt atcctacccc
 1981 aagcagtaca tttcacccat gggcaataat gggagctgta ccgtttggat ttctgctgac
 2041 ctgctgcatt tcttttatat aaatgtgact tttttttccc agaagttgat attaaact
 2101 attccagtct agtccttcta aactgttaat ttttaattaaa atgaagtact aatgactctt
 (SEQ ID NO:10)

Murine oxoglutarate dehydrogenase E430020N12

1 ggggggtggag ctgaacggga gacagggtact tgtggaaggc ttcaggacaa aatgtttcat
 61 ttaaggactt gtgctgctaa gttaaggcca ttgacagcct cccagactgt taagacattt
 121 tcacaaaaca aaccagcagc aattaggacg tttcaacaga ttcggtgcta ttctgcacct
 181 gtagctgctg aaccatttct tagtgggact agttcgaact atgtggagga aatgtactgt

241 gcctggttgg agaatcccaa aagtgtacat aagtcattggg acattttttt ccgaaacacc
 301 aatgctggag cccacccggg cactgcctac cagagccccc tttccctgag tcgaagctcc
 361 ctggctacca tggcccatgc acagtccttg gtggaagcac aacctaacgt cgacaaactc
 421 gtggaggacc acttggcggt gcagtctctc atcagggcat atcagatacg agggcaccat
 481 gtagcacagc tggaccccct ggggattttg gatgctgac tggactcctc cgtgcccgct
 541 gacattatct catccacaga caaacttggg ttctatggcc tacacgagtc tgaccttgac
 601 aagggtcttc acttaccac caccactttc atcgggggac aggagccagc acttcctctt
 661 cgggagatca tccgtcggct ggagatggcc tactgccagc acattggtgt ggagttcatg
 721 ttcattaatg atttggaaca atgccagtgg atccgacaga agtttgagac ccctggaatc
 781 atgcagttca ccaatgagga gaagcggacc ttgctggcca ggcttgtagc atccaccagg
 841 tttgaggagt tcctacagcg aaagtgggtc tcggagaagc gttttggtct ggaaggctgt
 901 gaggtgctga tccctgccct caagacaatc attgatattg caactcagat gaccctgaag
 961 ctgtcatgta tgtatgcaag gtggcagctg agtggagaaa caccttcac aaggatgttg
 1021 tagttgatct ggtgtgttat cgacgaaatg gccacaatga gatggacgaa cctatgttta
 1081 cacagccact catgtacaag cagatccgca agcagaagcc tgtactgcag aagtatgcag
 1141 aattgctagt ctcccagggt gtcgtcaatc agcctgagta cgaggaggaa atctccaagt
 1201 atgataagat ctgtgaggaa gcattttacca gatccaaaga tgagaagatc ttgcacatca
 1261 agcactggct ggattccccc tggcctggct ttttcaccct ggatggacag cccaggagca
 1321 tgacctgccc ctccactggc ctggaggagg atgtcttggt ccacattgga aagggtggcca
 1381 gctctgtacc tgtggagaac tttactatcc atggagggct gagccggatc ttgaagacct
 1441 gcagagagct tgtgacgaac cggactgtgg actgggccct ggcagagtac atggcatttg
 1501 gctcactgct gaaggaaggc atccatgtgc ggctgagtgg ccaggatgtg gagcggggca
 1561 ccttcagcca tcgccaccat gtgctccatg atcagaatgt tgacaaaaga acctgcatcc
 1621 ccatgaacca cctttggcca aatcaggccc cttacactgt atgcaacagc tcgctgtctg
 1681 agtacgggtgt cctgggcttt gagctgggct ttgccatggc tagccctaata gctctggttc
 1741 tctgggaggc ccagtttggt gacttcaaca acatggcaca gtgcatcatt gaccagttca
 1801 tctgcccagg acaggcaaag tgggtgcggc agaattggcat tgtgctcctg ctgcctcatg
 1861 gcatggaagg catgggtccc gagcattcct ctgaccgcc agagcgggtt ctgcatatgt
 1921 gcaatgatga cccatatgtc ctgcgtgact tgcaggaaga actctttgac atcaatcagc
 1981 tatatgactg caactggatt gttgtcagct gttccacccg tggcaacttc ttccatgtgc
 2041 tgcgacaaca gatcttgctg cccttcgta agccgttaata agtcttcact cccaaatccc

2101 tcttgcgcca ccgtgaggca agaactatct ttgacgatat gttgccagga acgcacttcc
 2161 agcgtgtgat ccagaaaaat ggacatgcag ctgaggaccc tcacaaagtc aagagacttc
 2221 tcttctgcac tgggaaggtg tactatgacc tcacccgaga gcgcaaagcc aggaacatga
 2281 aggaggaggt ggctattaca aggattgagc agctatcacc attccccctt gacctcctgt
 2341 tgaaagaggc tcagaagtat cccaatgctg agctggcctg gtgccaggaa gagcacaaga
 2401 accaaggcta ctatgactat gtcaagccaa gacttcgtac caccattgac cgtgctaagc
 2461 ctgtctggta tgctgtccga gaccggcag ctgctccagc cactggcaac aagaaaacac
 2521 acctgacaga gctgcagcgc tttctggaca cagcctttga cctggacgca ttcaagaaat
 2581 tctcttagat gctcctggag ttgatgaggc catggccccc atgtccatga cgctctttgc
 2641 ttctcaacta aagaatagtg cctcagcact gtccacacgt cccttcgctg tgccacacca
 2701 cccctgttct cataggaatt aagttgtcca ctgcagtgtc cagctgctcc ccggtcacat
 2761 gctgcccagc ctgtgccgac ttctctcagg ctgcacaccg ttcattggaga ccggaaggag
 2821 cagaataagg aaagggcccc tctcaggaca tcctagagaa ggaaggcagc tctggcccca
 2881 cccatgcccc cagtgcacac ctccagggtg ggaacagaac cctatgtggc ttcccagggt
 2941 actagcactc agccctcgtc acccatcaag tcgcagattc aaggccagga gtagtttcat
 3001 cttgctaggg ccaagctgag agctcatgga ggaactatag ctgccaggat ttgggagtca
 3061 tcaggatgtt gtgtgaatag agattgtcat ggggtattta gaggacttta gcagtgatgt
 3121 tagtctagcc ctgctaccct tcttgggttt gggctgtatg tgggaaactt accccagcta
 3181 ccacgcctgg agagcttggc tctgagtagc gcccagaagc tccattggct cccaacgcca
 3241 ggcactgctg cctcttggtc ctgctgcctc tgctctcctg acccctcccc agtcacttca
 3301 ttttctctgt tgttcccttg aacacacaga agctgttgac gaattctttt ttttgctgtg
 3361 ccaaggcagg tcaaaagcag atcagtggat aagagcaagt tgtcccaagg agccagctgt
 3421 ccttcctccc tcttttgacc tccactggga cacacctgat ttatttat ttggttaaaaa
 3481 aaaaaaggaa atgaaaaaag aacaaccacc tttgcattgc atcggttga ccataaact
 3541 aagttatcat ggtc (SEQ ID NO:11)

<Preparation of [cDNA + primer + aptamer + polymerase + PCR buffer composition] solution>

The cDNA, the primer set 1, the aptamer for Taq DNA polymerase and Taq DNA polymerase were mixed together to prepare a solution having the final concentrations of 5 μ M of the primer set 1, 5 μ M of the aptamer

for Taq DNA polymerase, 50 U/ μ l of Taq DNA polymerase, 0.1 M trehalose, 250 mM Tris-HCl (pH 8.3), 1.25 M KCl, 132.5 mM MgCl and 5 μ M of each dNTP.
<Spotting of DNA>

Each of the solutions prepared as described above was spotted on a sheet of 60MDP paper (Mishima Paper Co., Ltd., Japan) using a 96 pin-tool (Multi 96-multiblot replicator VP409, Bio Medical Science Inc., US) so that the position and the type of each spot could be identified, as shown in Fig. 15. Each spotting solution was applied at a rate of 1 μ l/spot.
<Amplification of DNA>

The spotted paper was dried at room temperature for at least 30 minutes. Thereafter, four 4 mm x 4 mm pieces were cut out from the 60MDP paper so that each piece contained the spotted area and then placed in a PCR microtube. The tube was added with 25 μ l of water, and PCR was run under the following conditions:

2 min at 94°C;

29 cycles of (1 min at 94°C, 1 min at 55°C, 75 sec at 68°C); and
then

15 min at 74°C.

After the reaction was completed, a proper amount of the reaction solution was removed from the tube and then subjected to 1% agarose gel electrophoresis. The results are shown in Fig. 16: Lane 1, malate dehydrogenase cDNA; Lane 2, isocitrate dehydrogenase (NADP) cDNA; Lane 3, isocitrate dehydrogenase (NAD) cDNA; Lane 4, oxoglutarate dehydrogenase cDNA; and left lane, size markers. It was demonstrated that fragments having desired lengths could be obtained for the four kinds of cDNA, respectively, and that PCR amplification was possible simply by adding water to the cDNA, primers, aptamer, polymerase and PCR buffer composition fixed on the 60MDP paper.

After the reaction was completed, a proper amount of the reaction solution was removed from the tube and then subjected to 1% agarose gel electrophoresis. The results are shown in 108. Bands observed at the

positions around 184 bp and 343 bp were considered to be the desired DNA fragments of exons 1 and 2, respectively. It was demonstrated that a desired fragment could be amplified by PCR from the template DNA by using the primers fixed on the 60MDP paper. Comparison was made between the samples with and without the aptamer for Taq DNA polymerase, and it was found that the reaction with the aptamer could inhibit the non-specific amplification.

[Example 4]

<Preparation of cDNA solution>

pFLC vector, into which the same murine malate dehydrogenase cDNA clone (Clone ID: 1500012M15, 1758 bp) as used in Example 3 had been cloned (Fig. 10), was dissolved in TE buffer (10 mM Tris-HCl, pH 8.0, 1 mM EDTA) to give a concentration of 0.1 µg/µl.

<Preparation of [cDNA + primer + reaction enhancer (spermidine) + polymerase + PCR buffer composition] solution>

The cDNA, the primer set 1, spermidine and Taq DNA polymerase were mixed together to prepare a solution having the final concentrations of 0.005 µg/µl of the cDNA, 5 µM of the primer set, 50 U/µl of Taq DNA polymerase, 100 mM of spermidine, 0.1 M of trehalose, 250 mM of Tris-HCl (pH 8.3), 1.25 M of KCl, 132.5 mM of MgCl and 5 µM of each dNTP.

<Spotting of DNA>

The solution prepared as described above was spotted on a sheet of 60MDP paper (Mishima Paper Co., Ltd., Japan) using a 96 pin-tool (Multi 96-multiblot replicator VP409, Bio Medical Science Inc., US) so that the position and the type of the spot could be identified, as shown in Fig. 15. The spotting solution was applied at a rate of 1 µl/spot.

<Amplification of DNA>

The spotted paper was dried at room temperature for at least 30 minutes. Thereafter, a 4 mm x 4 mm piece was cut out from the 60MDP paper so that the piece contained the spotted area and then placed in a PCR

microtube. The tube was added with 25 µl of water, and PCR was run under the following conditions:

2 min at 94°C;

29 cycles of (1 min at 94°C, 1 min at 55°C, 75 sec at 68°C); and then

15 min at 74°C.

After the reaction was completed, a proper amount of the reaction solution was removed from the tube and then subjected to 1% agarose gel electrophoresis. The results are shown in Fig. 17: Lanes 1 and 2, samples in which cDNA, primers, polymerase, spermidine (a polyamine) as reaction enhancer and PCR buffer composition were fixed on 60MDP paper; Lane 3, a sample fixing the same solution as those of Lanes 1 and 2 except that spermidine was eliminated. It was demonstrated that for each of the samples, the fixed DNA could be amplified by adding water and that in the samples on which a reaction enhancer was also fixed, the amplification reaction was enhanced.

All publications, patents and patent applications cited herein are incorporated herein by reference in their entirety.

Industrial Applicability

The support according to the present invention can be used for storage and distribution of enzymes. The support is also applicable to printed materials and reagent kits.

Free Text of Sequence Listing

SEQ ID NO:1 shows the nucleotide sequence of primer -21M13.

SEQ ID NO:2 shows the nucleotide sequence of primer 1233-Rv.

SEQ ID NO:3 shows the nucleotide sequence of cDNA for murine malate dehydrogenase.

SEQ ID NO:4 shows the nucleotide sequence of primer HsLH1F.

SEQ ID NO:5 shows the nucleotide sequence of primer HsLH1R.

SEQ ID NO:6 shows the nucleotide sequence of primer HsLH2F.

SEQ ID NO:7 shows the nucleotide sequence of primer HsLH2R.

SEQ ID NO:8 shows the nucleotide sequence of an aptamer for Taq DNA polymerase.

SEQ ID NO:9 shows the nucleotide sequence of cDNA for murine isocitrate dehydrogenase (NADP).

SEQ ID NO:10 shows the nucleotide sequence of cDNA for murine isocitrate dehydrogenase (NAD).

SEQ ID NO:11 shows the nucleotide sequence of cDNA for murine oxoglutarate dehydrogenase.